

EXPLORING CIRCULAR BACTERIOCINS: NEW APPROACHES FOR PRODUCTION AND ENGINEERING



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BACTERIOCINS

HEAT-STABLE (<10 kDa)

Class I – Modified

- Lanthipeptides

- Circular peptides

- LAPs

- Sactibiotics

- Glycocins

- Lasso peptides

Class II – Unmodified

- Pediocin-like

- Two-peptides

- Leaderless

- Non pediocin-like single-peptides

THERMO-LABILE (>10 kDa)

Class III

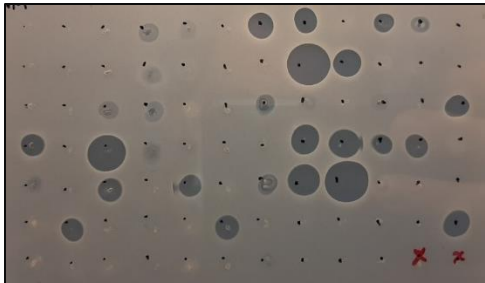
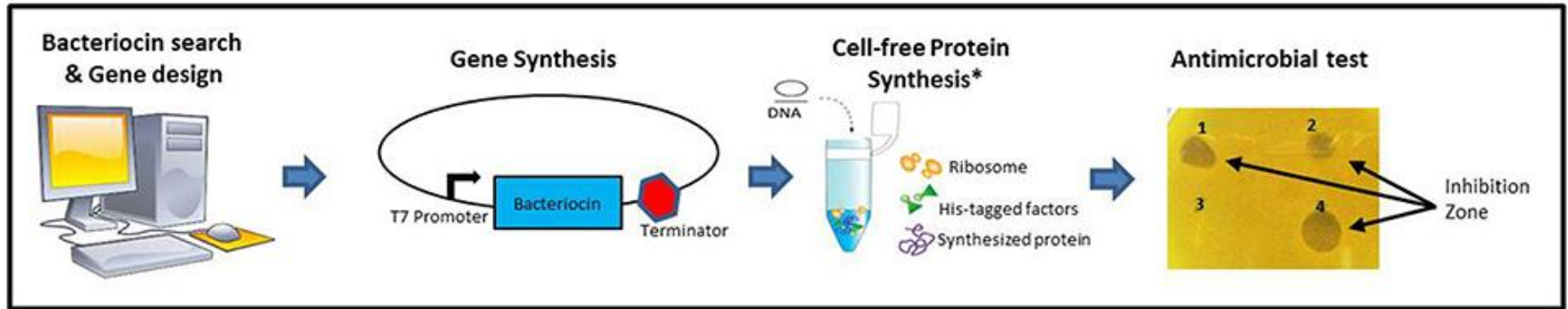
- Bacteriolysins

- Non-lytic



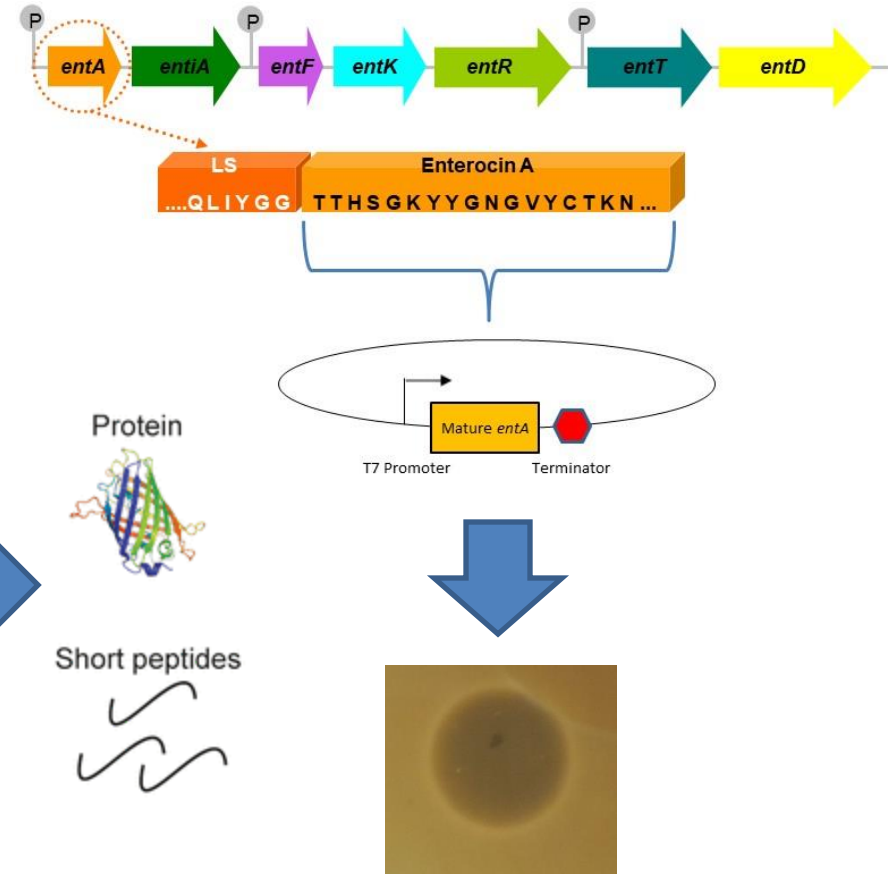
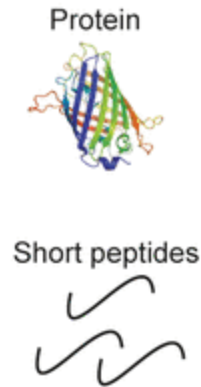
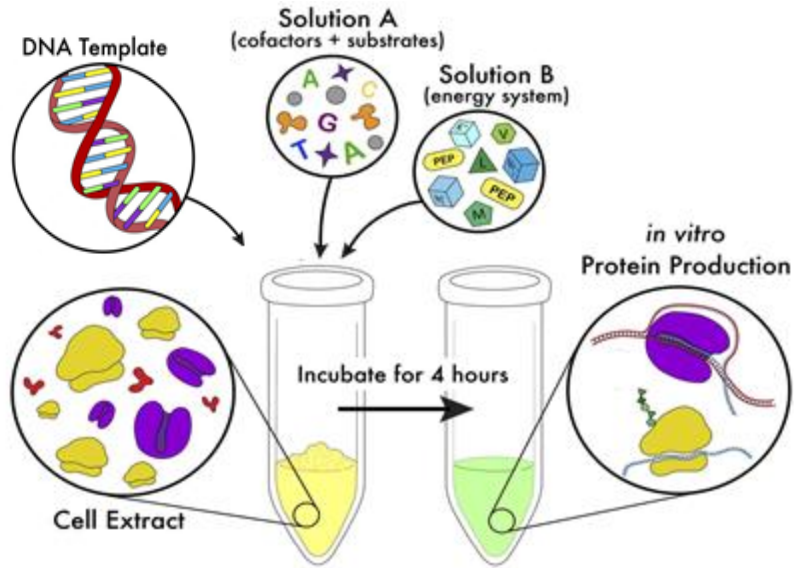
PARAGEN 1.0: A Standardized Synthetic Gene Library for Fast Cell-Free Bacteriocin Synthesis

Philippe Gabant* and Juan Borrero*

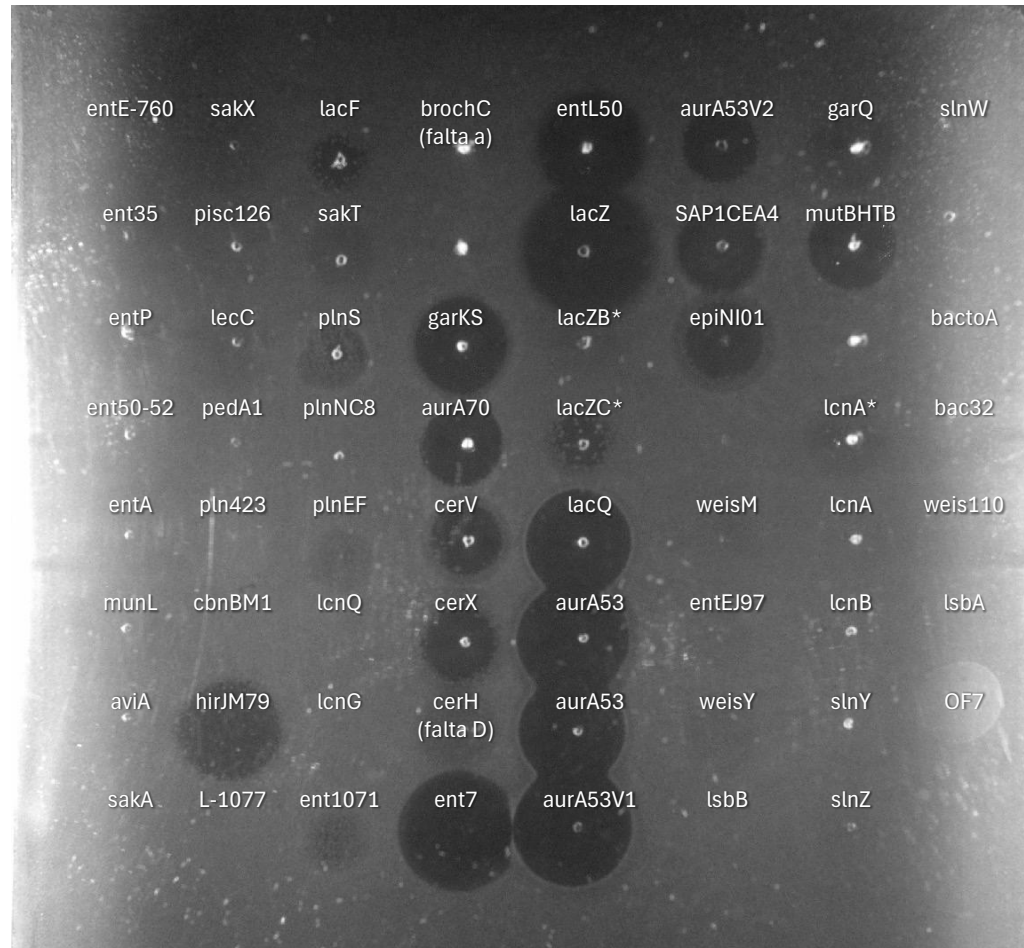


~170 bacteriocins

➤ Cell-free *in vitro* Protein Synthesis



Activity Vs *Streptococcus suis*



BACTERIOCINS

HEAT-STABLE (<10 kDa)

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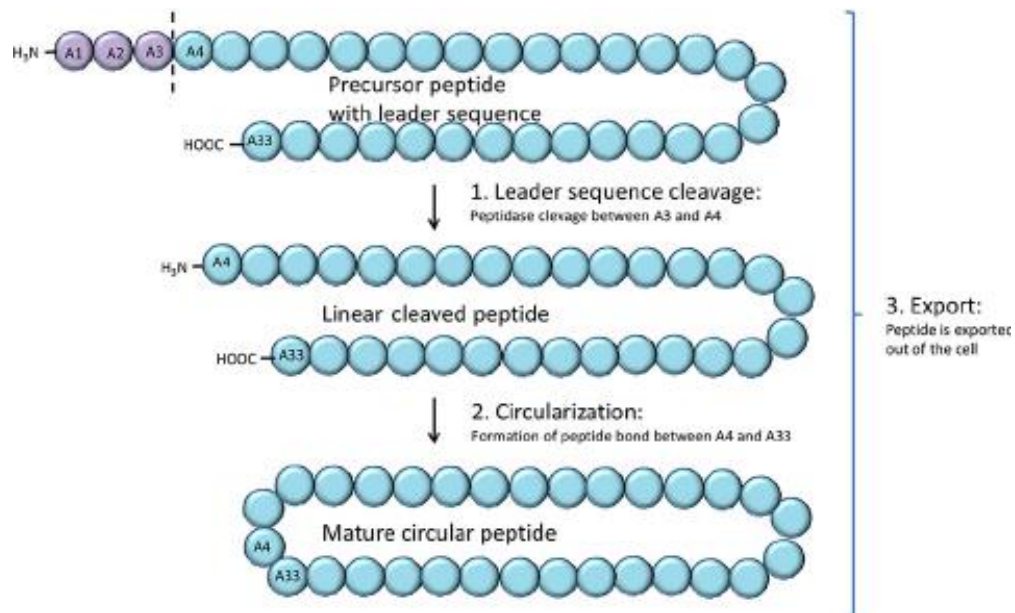
- Non pediocin-like single-peptides

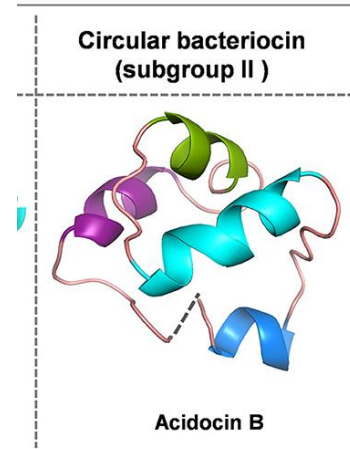
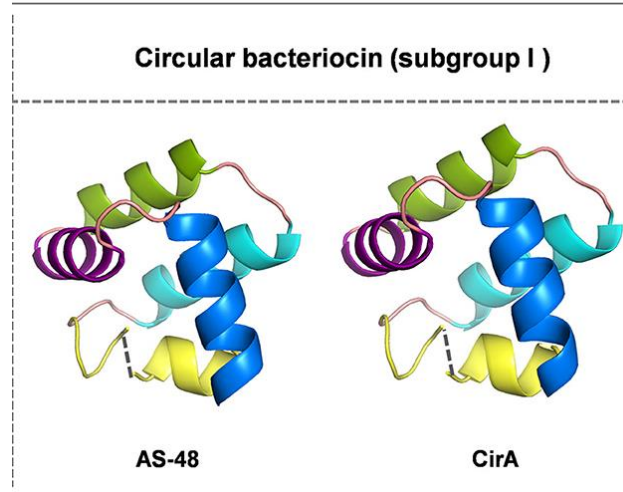
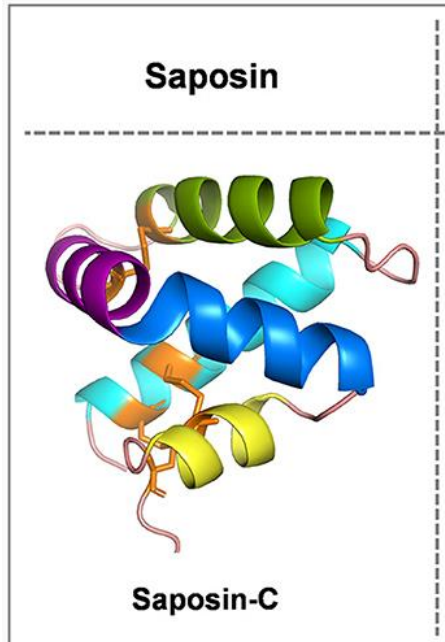
THERMO-LABILE (>10 kDa)

Class III

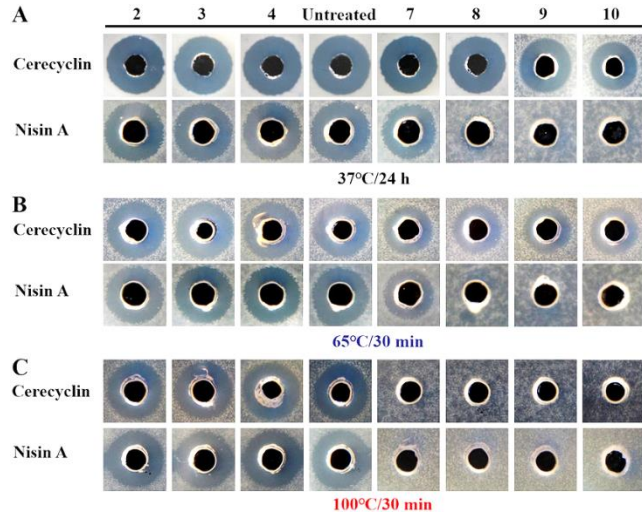
- Bacteriolysins

- Non-lytic





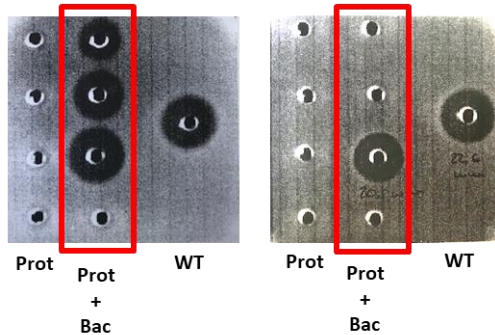
Xin et al., 2020, mSystems.



Proteases

garvicin ML

enterocin A

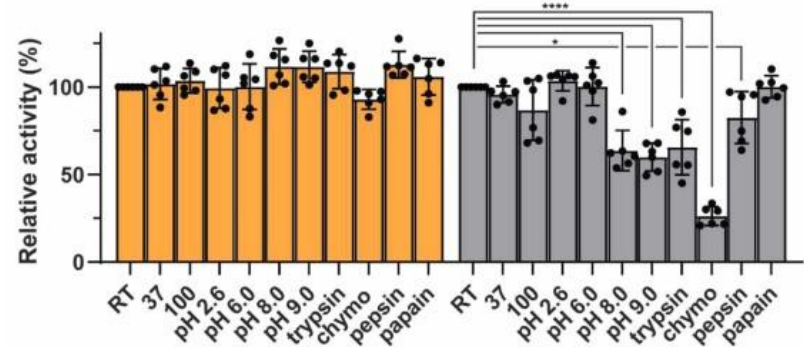
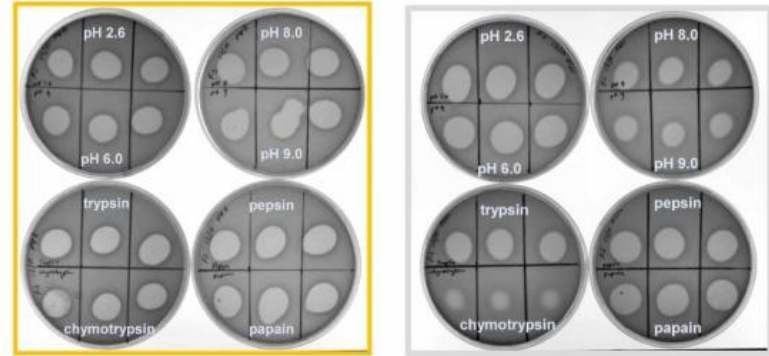


LINEAR Vs CIRCULAR

A. Antibacterial activity

Ent53B

nisin



Wang et al., 2023, Peptides.

RESEARCH ARTICLE

Open Access

Bioinformatic prospecting and phylogenetic analysis reveals 94 undescribed circular bacteriocins and key motifs

Ben Vezina, Bernd H. A. Rehm and Andrew T. Smith*



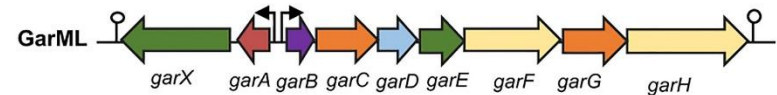
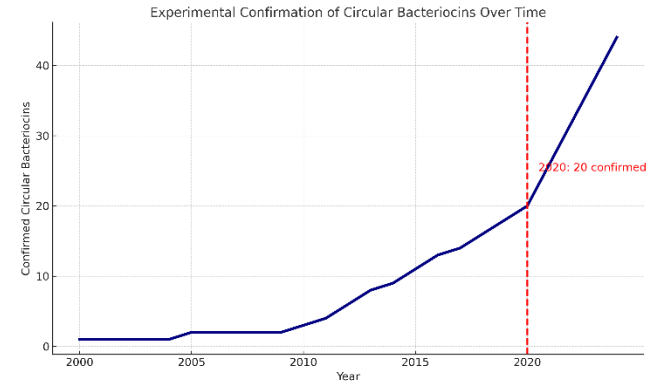
RESEARCH ARTICLE
 Applied and Environmental Science



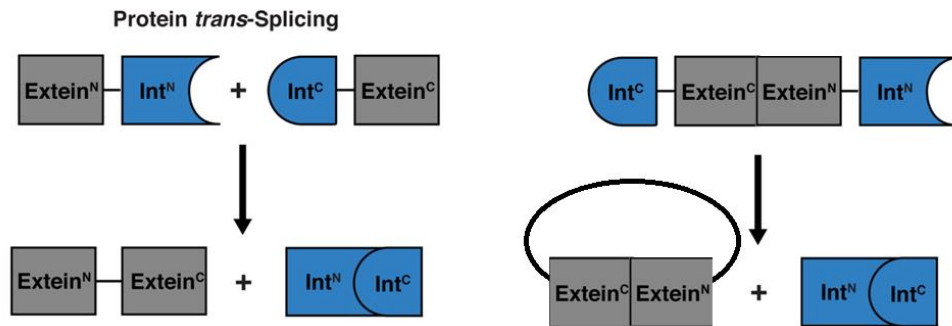
In Silico Analysis Highlights the Diversity and Novelty of Circular Bacteriocins in Sequenced Microbial Genomes

Bingyue Xin,^{a,b,c} Hualin Liu,^a Jinshui Zheng,^{a,d} Chuanshual Xie,^a Ying Gao,^a Dadong Dai,^a Donghai Peng,^a Lifang Ruan,^a Huanchun Chen,^c Ming Sun^a

Experimentally confirmed circular bacteriocins



Inteins



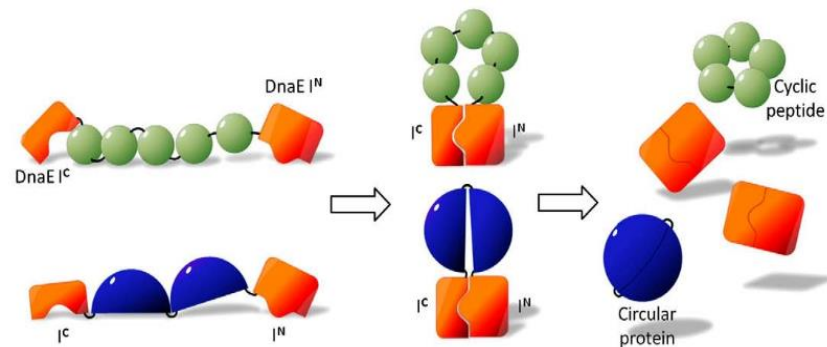
RESEARCH ARTICLE

Production of cyclic peptides and proteins *in vivo*

Charles P. Scott, Ernesto Abel-Santos, Mark Wall, Daphne C. Wahnou, and Stephen J. Benkovic

PNAS November 23, 1999 96 (24) 13638-13643; <https://doi.org/10.1073/pnas.96.24.13638>

Contributed by Stephen J. Benkovic



GARVICIN ML

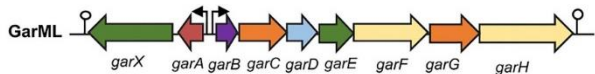
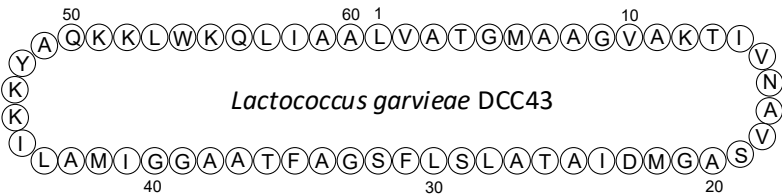
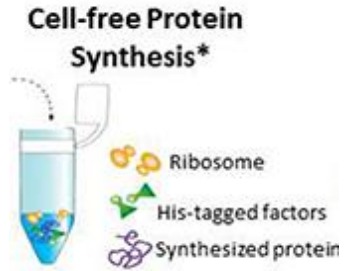
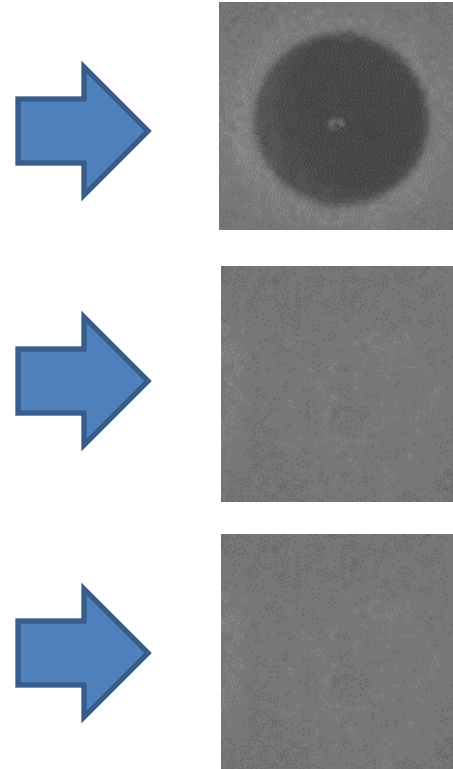
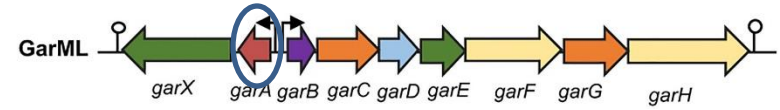
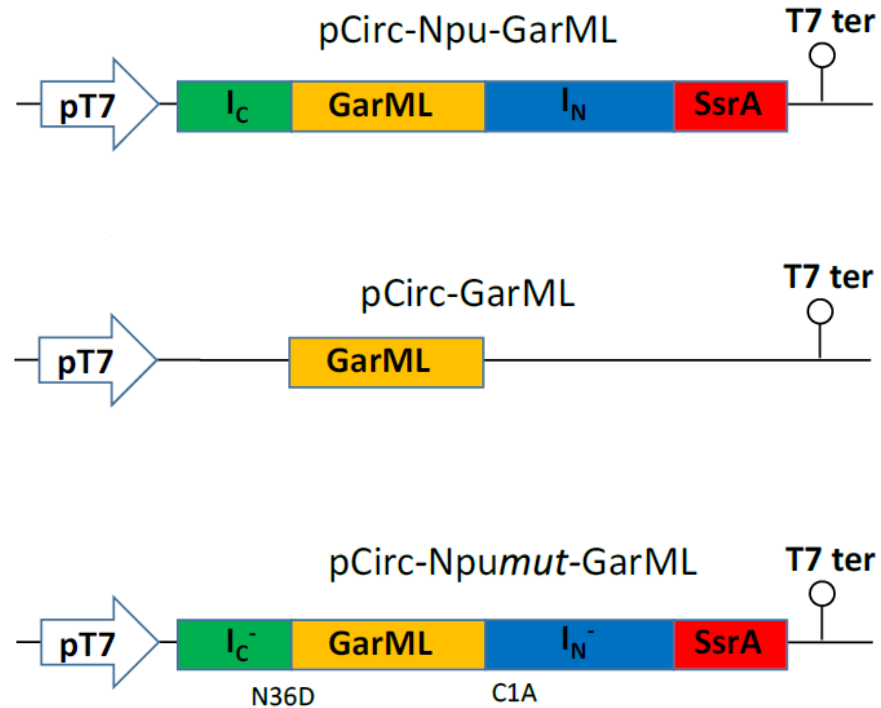


TABLE 1. Antimicrobial activities and inhibitory spectrum of fractions generated from the purification of garvicin ML produced by *L. garvieae* DCC43^a

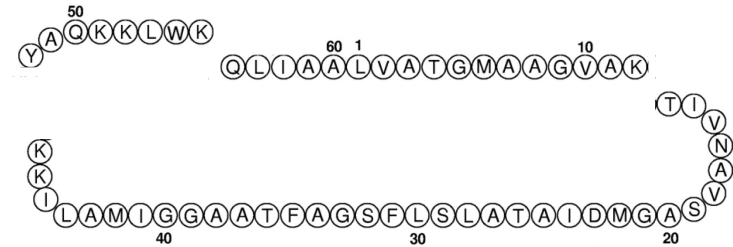
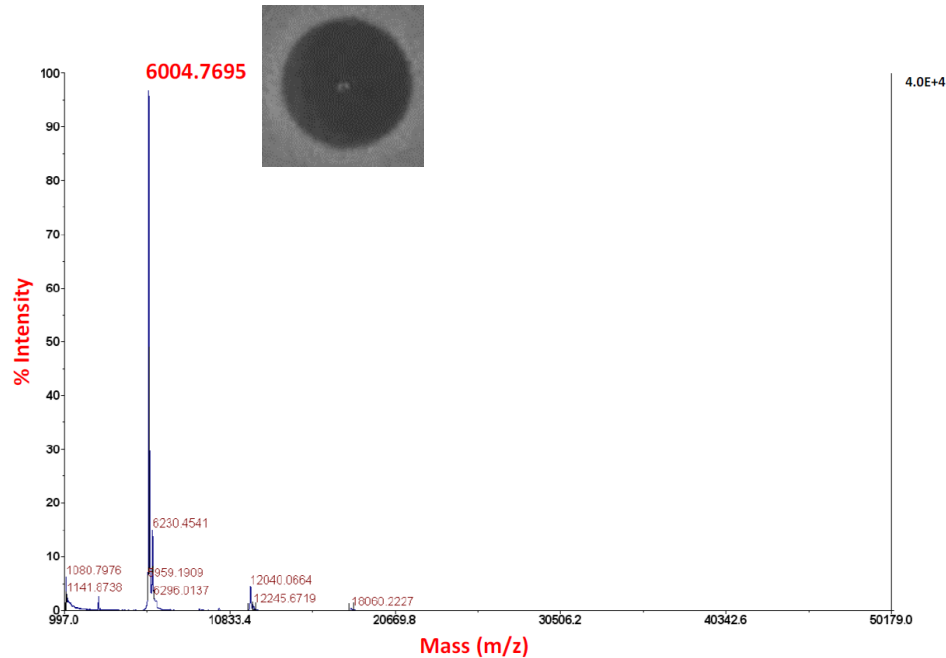
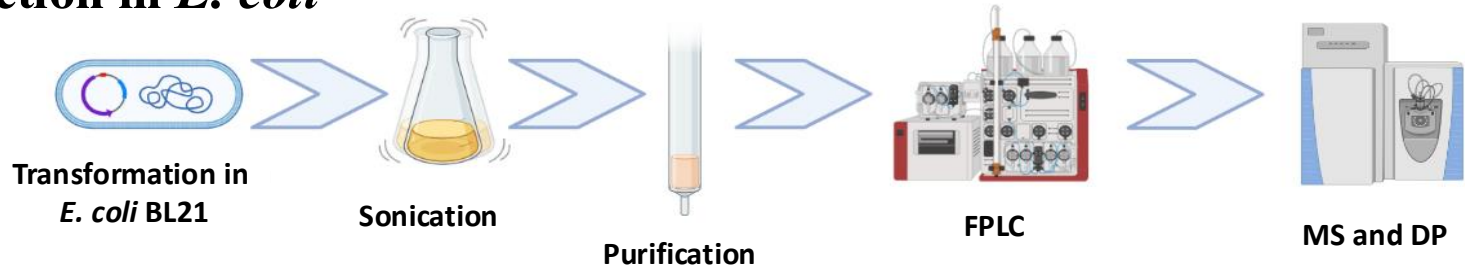
Indicator strain	Source ^b	Halo of inhibition (mm ²) using indicated garvicin ML fraction ^c					
		SN	AS	GF	SE	OE	RP
<i>Lactobacillus reuteri</i> 20016	DSM	— ^d	—	—	51	369	378
<i>Lactobacillus helveticus</i> 15009	ATCC	—	—	22	26	129	492
<i>Lactobacillus curvatus</i> 2739	NCFB	—	—	—	—	221	177
<i>Lactobacillus casei</i> 334	ATCC	83	215	153	191	1,048	1,174
<i>Lactobacillus acidophilus</i> 4356	ATCC	—	—	—	25	149	236
<i>Lactobacillus sakei</i> 2714	NCFB	94	204	163	249	1,445	1,445
<i>Lactococcus lactis</i> BB24	FVM	97	186	150	183	941	928
<i>Lactococcus lactis</i> NZ9000	NIZO	—	69	37	77	163	163
<i>Lactococcus lactis</i> DPC5598	DPC	163	98	113	141	1,075	1,541
<i>Lactococcus garvieae</i> 5274	CECT	—	67	71	94	684	617
<i>Lactococcus garvieae</i> 5806	CECT	334	482	413	561	2,807	2,807
<i>Lactococcus garvieae</i> 5807	CECT	291	438	441	529	2,603	2,705
<i>Lactococcus raffinolactis</i> 988	CECT	—	—	—	10	228	228
<i>Pediococcus acidilactici</i> 347	FVM	94	193	153	235	1,398	1,398
<i>Pediococcus pentosaceus</i> FBB61	TNO	21	144	108	193	1,131	1,131
<i>Enterococcus faecium</i> P13	FVM	65	201	160	232	1,217	1,291
<i>Enterococcus faecium</i> L50	FVM	—	63	47	83	574	719
<i>Enterococcus faecalis</i> DBH18	FVM	—	—	—	—	25	25
<i>Enterococcus faecalis</i> P4	IFR	—	—	—	—	86	123
<i>Propionibacterium</i> sp. P6	NCDO	62	119	111	130	684	190
<i>Propionibacterium acidipropionici</i> 563	NCDO	75	153	105	132	662	864
<i>Clostridium tyrobutyricum</i> 3.5 CT	TNO	55	124	83	124	719	651
<i>Clostridium tyrobutyricum</i> 1754	NCDO	58	129	105	126	802	839
<i>Clostridium perfringens</i> 376	CECT	—	45	29	31	351	352
<i>Clostridium botulinum</i> 551	CECT	—	59	174	227	1,232	574
<i>Listeria monocytogenes</i> 4032	CECT	—	22	215	289	1,574	662
<i>Listeria ivanovii</i> 913	CECT	—	139	273	353	1,978	1,020
<i>Listeria seeligeri</i> 917	CECT	34	87	212	292	1,509	684
<i>Listeria grayi</i> 931	CECT	—	127	135	191	1,075	742
<i>Listeria welshimeri</i> 919	CECT	—	—	158	258	1,291	452
<i>Brochothrix thermosphacta</i> 847	CECT	—	—	—	31	255	283
<i>Pseudomonas fluorescens</i> 378	CECT	—	—	—	—	—	—
<i>Escherichia coli</i> JM109	Invitrogen	—	—	—	—	—	—
<i>Escherichia coli</i> MC1000	NIZO	—	—	—	—	—	—
<i>Salmonella paratyphi</i> 554	CECT	—	—	—	—	—	—
<i>Salmonella</i> Typhimurium 443	CECT	—	—	—	—	—	—
<i>Salmonella enteritidis</i> 4396	CECT	—	—	—	—	—	—
<i>Streptococcus pneumoniae</i> FQ26	HRC	69	156	100	110	599	590
<i>Streptococcus pneumoniae</i> 67620	HRC	65	135	94	128	707	596
<i>Streptococcus pneumoniae</i> 15 M-1047	HRC	58	128	96	139	790	790



➤ Cell-free *in vitro* Protein Synthesis



➤ Production in *E. coli*



Skyline			
Sequence	E-1+2	E-3	
QLIAALVATGMAAGVAK	Si	Si(MS/MS)	
QLIAALVATGMAAGVAK	Si (MS/MS)	Si(MS/MS)	
IAALVATGMAAGVAK	No	Si(MS/MS)	
LVATGMAAGVAK	No	Si(MS/MS)	
LVATGMAAGVAK	Si(MS/MS)	No	
YAQKKLWK	Si(MS/MS)	Si(MS/MS)	
TIVNAVSAGMDIATLSLFSGAFTAAAGGIMALIKK	No	Si (MS/MS)	
TIVNAVSAGMDIATLSLFSGAFTAAAGGIMALIK	No	Si(MS/MS)	
TIVNAVSAGMDIATLSLFSGAFTAAAGGIMALIK	No	Si (MS/MS)	
TIVNAVSAGMDIATLSLFSGAFTAAAGGIMALIK	No	Si (MS/MS)	

Characterized Circular bacteriocins

Bacteriocin	Producer	Plasmid	Activity against <i>L. lactis</i> IL1403*
Amylocyclin	<i>Bacillus amyloliquefaciens</i> FZB42	pCirc-Npu-Alc	+
Enterocin AS-48	<i>Enterococcus faecalis</i> S-48	pCirc-Npu-EntAS48	+
Carnocyclin A	<i>Carnobacterium maltaromaticum</i> UAL307	pCirc-Npu-CarA	+
Circularin A	<i>Clostridium beijerinckii</i> ATCC 25752	pCirc-Npu-CirA	+
Enterocin NKR-5-3B	<i>Enterococcus faecium</i> NKR-5-3	pCirc-Npu-NKR_5_3B	+
Garvicin ML	<i>Lactococcus garvieae</i> DCC43	pCirc-Npu-GarML	+
Leucocyclin Q	<i>Leuconostoc mesenteroides</i> TK41401	pCirc-Npu-LeuQ	+
Uberolysin A	<i>Streptococcus uberis</i> 42	pCirc-Npu-UberA	–
Butyrivibriocin AR10	<i>Butyrivibrio fibrisolvens</i> AR10	pCirc-Npu-ButAR10	–
Paracyclin P	<i>Lactobacillus paracesei</i> JCM 8130/ DSM 5622	pCirc-Npu-ParP	+
Gassericin A	<i>Lactobacillus gasseri</i> LA39	pCirc-Npu-GasA	+
Plantaricyclin A	<i>Lactobacillus plantarum</i> NI326	pCirc-Npu-PlcA	+
Cerecyclin	<i>Bacillus</i> sp. Xin1	pCirc-Npu-Cer	–
Bacteriocin 3688STDY6124959	<i>Staphylococcus aureus</i> 3688STDY6124959	pCirc-Npu-3688STDY	+
Bacteriocin BCW 2997	<i>Listeria monocytogenes</i> BCW 2997	pCirc-Npu-BCW_2,997	+
Bacteriocin CF11	<i>Clavibacter michiganensis</i> CF11	pCirc-Npu-CF11	–
Bacteriocin NBRC 15376	<i>Paenibacillus chondroitinus</i> NBRC 15376	pCirc-Npu-NBRC_15,376	–
Bacteriocin YS111	<i>Streptococcus suis</i> YS111	pCirc-Npu-YS111	+
Bacteriocin DSM 15102	<i>Garcilla nitratireducens</i> DSM 15102	pCirc-Npu-DSM_15,102	+
Bacteriocin AFS089278	<i>Bacillus toyonensis</i> AFS089278	pCirc-Npu-AFS089278	–
Bacteriocin TD3	<i>Bacillus vallismortis</i> TD3	pCirc-Npu-TD3	–
Bacteriocin NRRL B-24287	<i>Streptomyces pathocidini</i> NRRL B-24287	pCirc-Npu-NRLL_B_24,287	–
Bacteriocin AK22	<i>Alkalibacterium</i> AK22	pCirc-Npu-AK22	–
Bacteriocin 15,828	<i>Gemella cuniculi</i> DSM 15828	pCirc-Npu-15,828	–
Bacteriocin NCTC 12958	<i>Streptococcus thermophilus</i> NCTC 12958	pCirc-Npu-NCTC_12,958	+
Bacteriocin UoS2029	<i>Streptococcus pneumoniae</i> UoS2029	pCirc-Npu-UoS2029	–

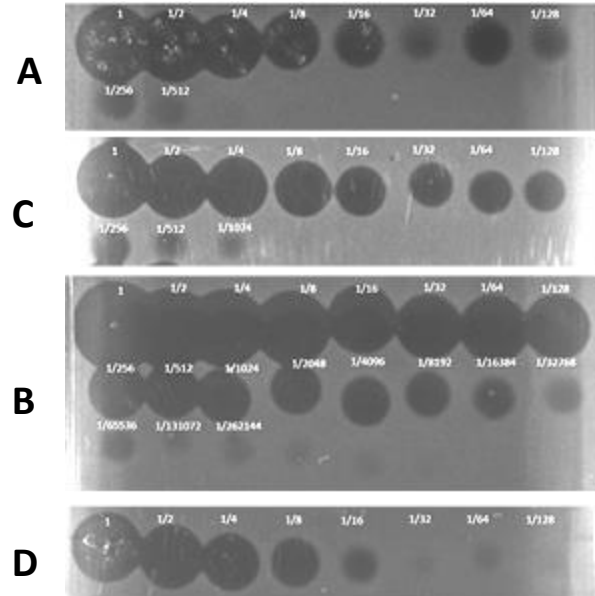
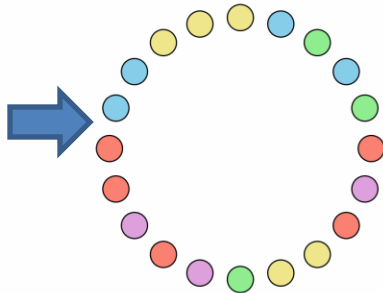
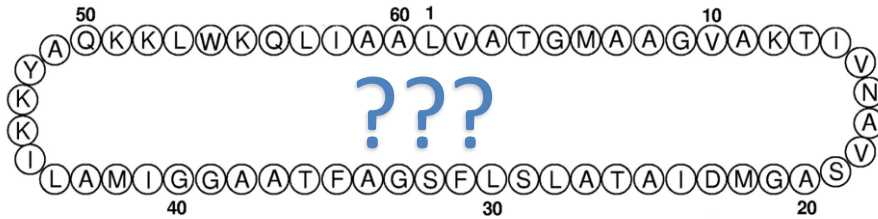
*Samples showing a clear halo of inhibition (+) or no halo of inhibition (–).

Non Characterized Circular bacteriocins

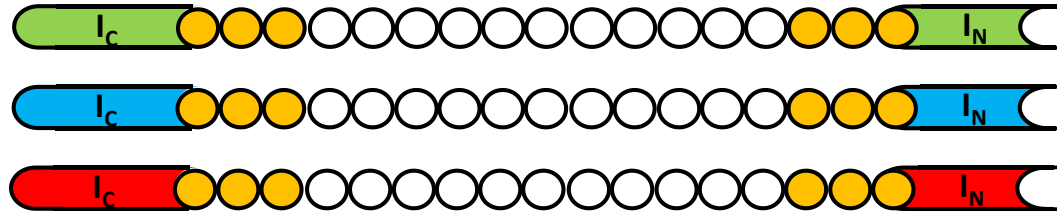
IMPROVEMENTS OF THE SIML SYSTEM



TESTING DIFFERENT CIRCULARIZATION SITES



IMPROVEMENTS OF THE SIML SYSTEM



TESTING DIFFERENT INTEINS

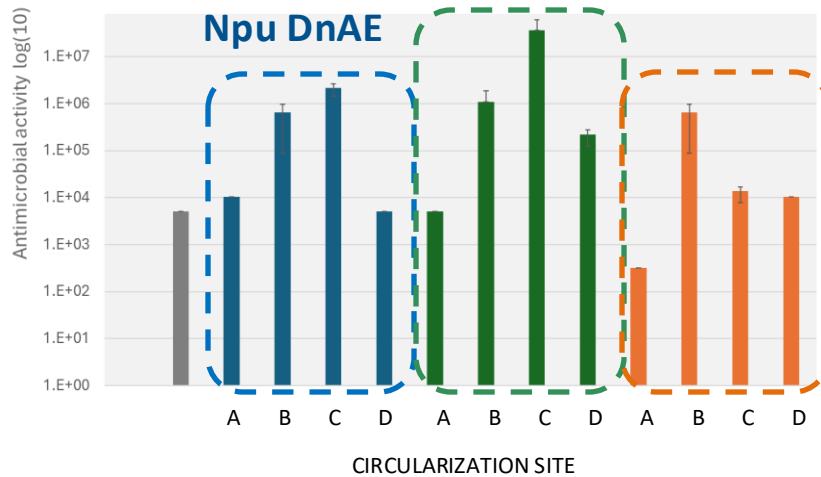


Table 1. Antimicrobial activity of variants of circular bacteriocins against selected indicator bacteria.

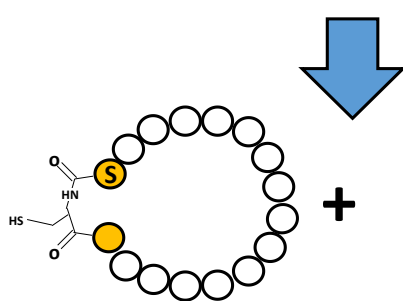
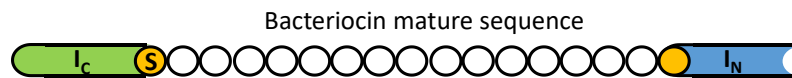
	<i>P. damnosus</i>	<i>L. garvieae</i>	<i>E. faecium</i>	<i>L. monocytogenes</i>	<i>S. aureus</i>	<i>S. suis</i>	<i>B. cereus</i>	<i>E. rhusiopathiae</i>	<i>S. agalactiae</i>	<i>P. larvae</i>	<i>C. perfringens</i>	<i>E. coli</i>
BAC1	+++	+++	+++	+++	-	+++	+++	+++	+	+++	++	-
BAC2	+++	+++	+++	+++	+	+++	+++	+++	-	+++	++	-
BAC3	++	+	+	+++	-	+	+++	-	-	+++	-	-
BAC4	++	++	-	++	-	-	++	+	-	+	-	-
BAC5	++	+	-	+	-	-	++	-	-	++	-	-
BAC6	++	++	+	+	-	+	++	++	-	+	+	-
BAC7	+				-	-	-	-	-	-	-	-
BAC8	+				-	-	-	-	-	-	-	-
BAC9	++				-	-	+	-	-	-	-	-
BAC10	+++				-	+++	-	+	-	-	+	-
BAC11	+++				-	+	-	++	-	++	-	-
BAC12	++				-	-	-	-	-	-	-	-
BAC13	-				-	-	-	+	-	-	-	-
BAC14	++				-	-	-	+++	-	++	-	-
BAC15	+				-	-	-	-	-	-	-	-
BAC16	+++				-	+	++	-	+	+	+	-
BAC17	+++				-	+	++	-	-	+	-	-
BAC18	+++				-	+	++	-	+	++	+	-
BAC19	+++				-	+	++	-	-	+	-	-
BAC20	+				-	+	-	-	-	-	-	-
BAC21	++				-	++	-	+	-	-	+	-
BAC22	++	++	+	++	-	+	-	-	-	-	-	-
BAC23	++	++	+	+	-	+	-	++	-	-	+	-
BAC24	++	-	-	+	-	-	-	-	-	-	-	-
BAC25	+++	+++	+++	+	-	++	-	-	-	-	++	-
BAC26	+++	+	-	++	-	+	-	-	-	-	-	-
BAC27	-	-	-	-	-	-	-	-	-	-	-	-



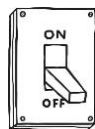
MAGADAN MAMMOTH RECOVERED FROM THE PERMAFROST 1977

Antimicrobial activity performed by a spot-on-agar-test (SOAT) and calculated as the diameter of the zone of inhibition, -: no inhibition, +: <5 mm, ++: 5–10 mm, +++: >10 mm.

SWITCHABLE INTEINS FOR CONDITIONAL PROTEIN SPLICING

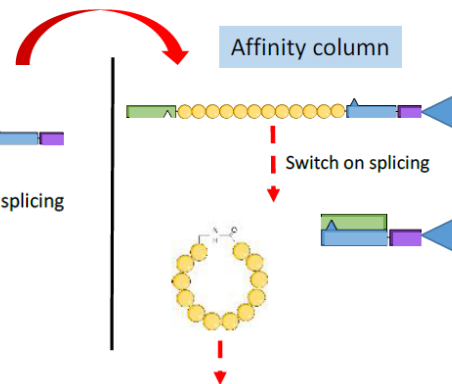
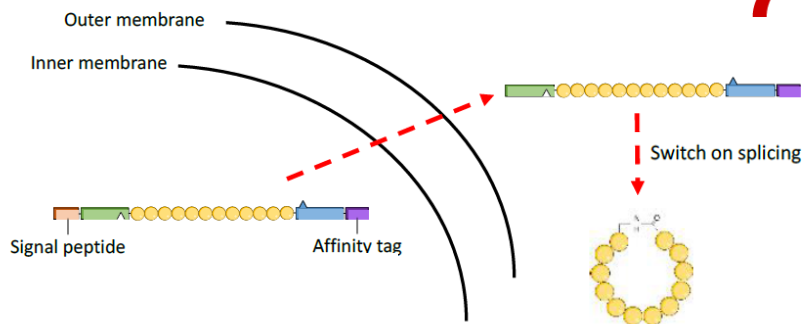


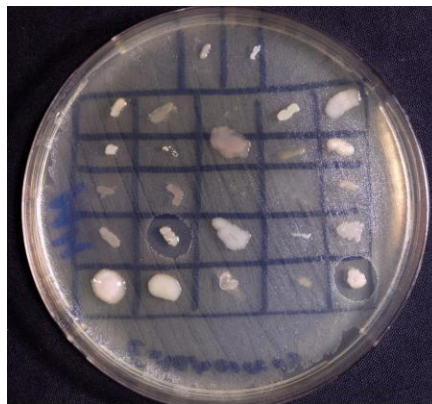
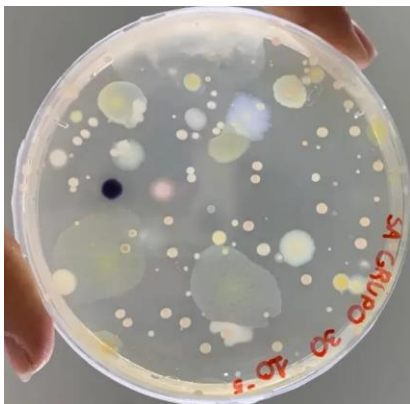
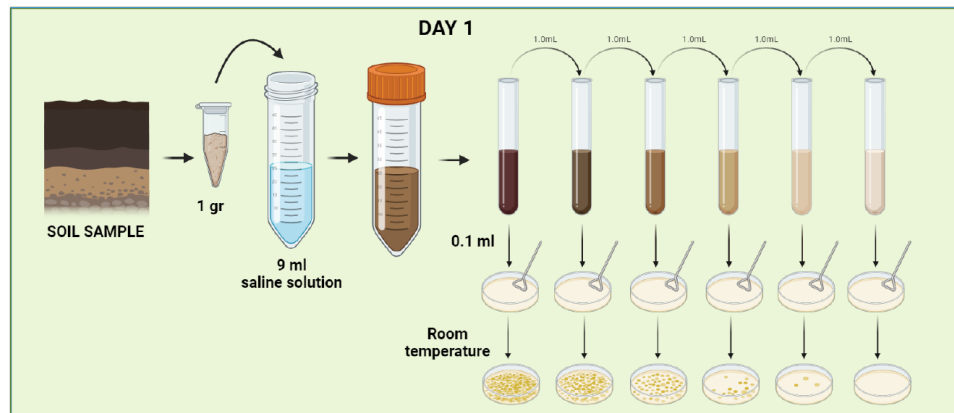
Circular bacteriocin



INCREASE PROTEIN YIELDS

IMPROVE PURIFICATION PROCESS





tiny  earth

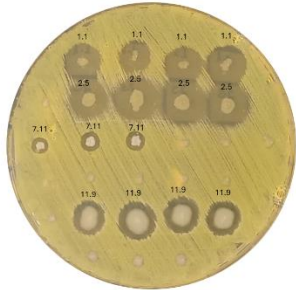
 Small World Initiative
crowdsourcing antibiotic discovery

MicroMundo@UCM

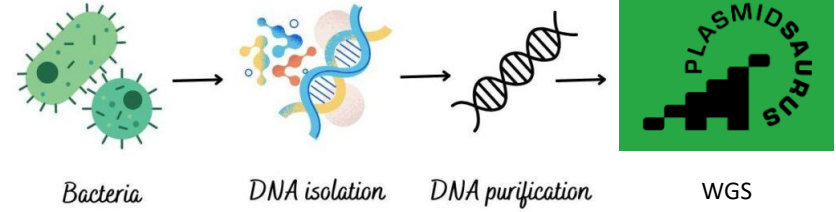


Gram (+)
Kocuria rhizophila

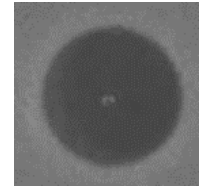
Gram (-)
E. coli Nissle 1917



1.1. *Paenibacillus polymyxa*
2.5. *Bacillus pumilus*
7.11. *Staphylococcus epidermidis*
11.9. *Bacillus cereus*



BAGEL4



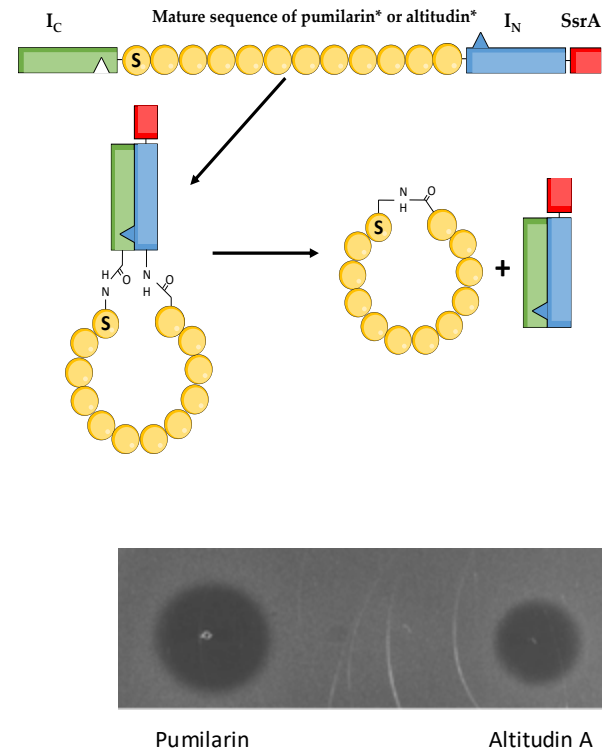
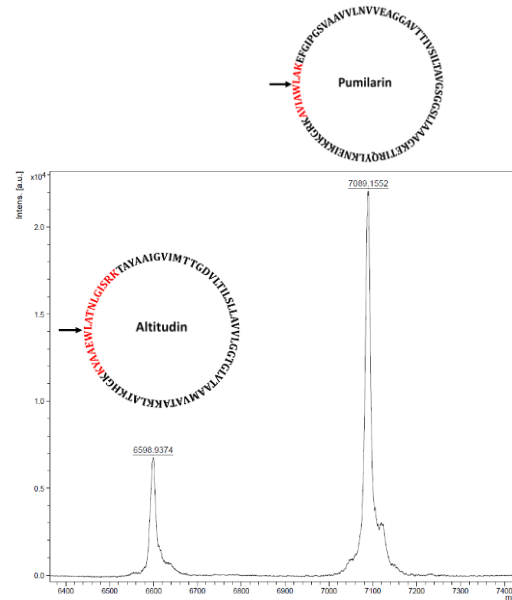
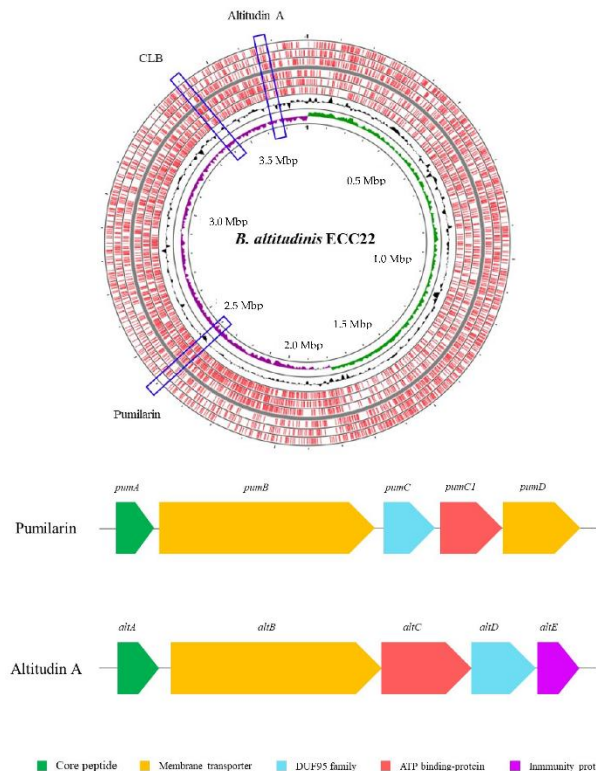
tiny earth

Small World Initiative
crowdsourcing antibiotic discovery

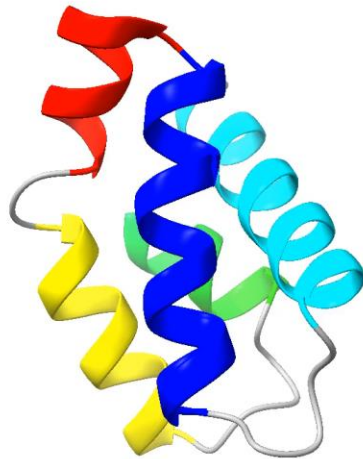
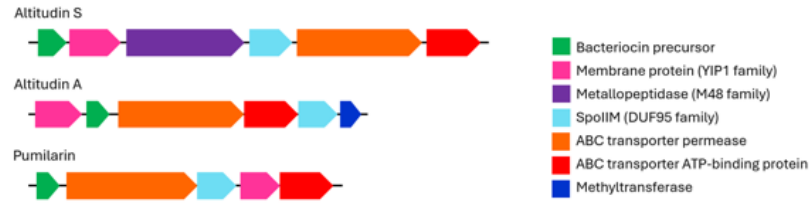
MicroMundo@UCM

Production of Pumilarin and a Novel Circular Bacteriocin, Altitudin A, by *Bacillus altitudinis* ECC22, a Soil-Derived Bacteriocin Producer

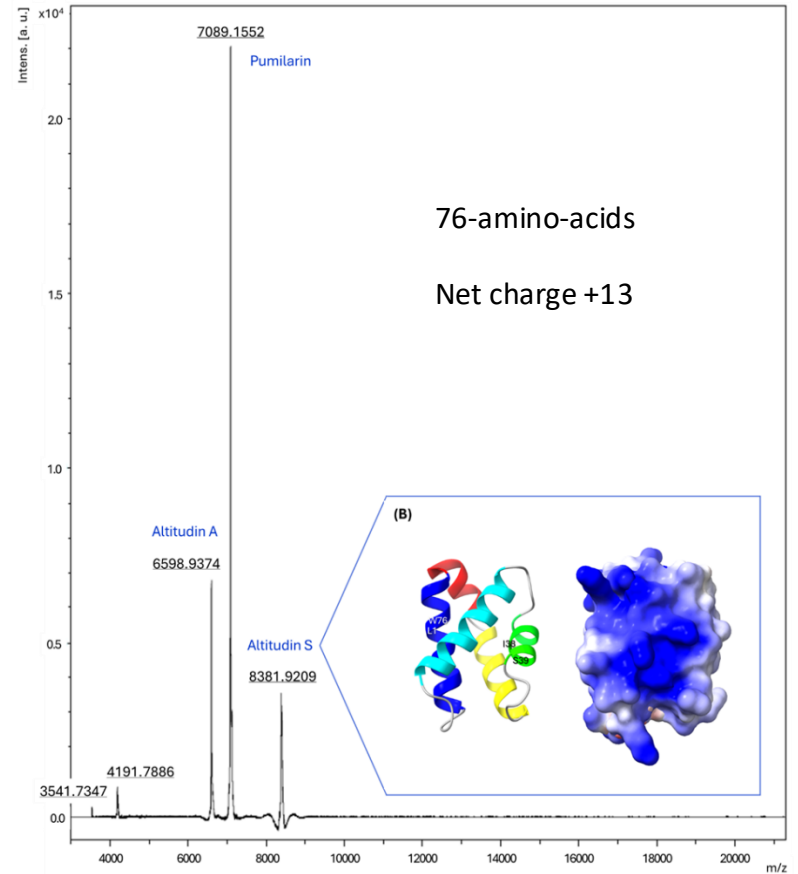
by Irene Lafuente ¹ ✉, Ester Sevillano ¹ ✉, Nuria Peña ¹ ✉, Alicia Cuartero ² ✉, Pablo E. Hernández ¹ ✉, Luis M. Cintas ¹ ✉, Estefanía Muñoz-Atienza ^{1,*} ✉ and Juan Borrero ^{1,*} ✉



ALTITUDIN S



(A)



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